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EXAMINER

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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

RESPONSE TO AMENDMENT

The amendment filed 9-18-08 has been entered into the record. Claims 1-13, 12-22, 25-32, 34 and 36-37 have been cancelled. Claims 14, 23, 24, 33, 35, 38-57 are pending. Claims 14, 23, 24, 33, 35, 38, 39, 40, 50 and 54 are under examination.

The text of Title 35 of the U.S. Code not reiterated herein can be found in the previous office action.

Election/Restrictions

Newly submitted claims 41-49, 51-53 and 55-57 are directed to an invention that is independent or distinct from the invention originally claimed for the following reasons: the claims are drawn to a complex of the antibody and its specific binding partner beta amyloid in cerebrospinal fluid. As such, the composition is patentably distinct from the antibody *per se* as the composition functions in a materially different manner because it cannot bind soluble beta amyloid since the antibody is already bound to the antigen present in the cerebrospinal fluid.

Since applicant has received an action on the merits for the originally presented invention, this invention has been constructively elected by original presentation for prosecution on the merits. Accordingly, claims 41-49, 51-53 and 55-57 are withdrawn from consideration as being directed to a non-elected invention. See 37 CFR 1.142(b) and MPEP § 821.03.

Rejections Withdrawn

All previous art rejections are withdrawn in favor of the new grounds of rejection set forth below.

Rejections Maintained

The rejection of claims 39 and 40 are rejected under 35 U.S.C. 112, first paragraph, as failing to comply with the written description requirement is maintained for reasons made of record and herein.

The claims are drawn to neutralizing antibodies that inhibit neurotoxicity and bind soluble beta amyloid. It was well established in the art at the time of the invention and admitted by Applicants in the specification that neurotoxicity was the result of fibrillar or aggregated beta amyloid and as such, an antibody that binds soluble beta amyloid cannot neutralize it's neurotoxicity because it is not neurotoxic in the first place. The specification does not teach that soluble amyloid is neurotoxic and as such conception of neurotoxicity associated with soluble amyloid neutralization thereof is deemed a new concept not present in the specification at the time of filing.

Applicants point to specific pages of the specification that teach that the therapeutic benefit of the antibody is to sequester the soluble amyloid beta to possibly inhibit the formation of aggregates. This is not neutralization of neurotoxicity of soluble amyloid, which is specifically claimed. Applicants point to pages 10-11 which again do not establish that the claimed soluble beta amyloid is neurotoxic. Nothing in the passage indicates that the claimed soluble beta amyloid is neurotoxic and that binding it neutralizes it's neurotoxicity. The art does not recognize that the amyloid of the claims is neurotoxic. The neurotoxicity of the art is associated with deposited/aggregated/fibrillar amyloid beta which is not soluble.

New Rejections Based on Amendment

Claims 14, 24, 39 and new claim 50 stand are rejected under 35 U.S.C. 103(a) as being unpatentable over Saido et al (The Journal of Biochemistry, 269(21):15253-15257, 1994, Fee-based IDS Nov 16, 2001) in view of Takeda Chemical Industries Ltd., (EP 0 683 234 A1, published November 22, 1995, reference AC on PTOL-1449 filed 12 October 1999), Vigo-Pelfrey et al (Journal of Neurochemistry, 61:1965-1968, 1993), Goding

(Monoclonal Antibodies, Academic Press Inc., London 1983, pages 56-97) and Queen et al (US Patent No, 6,180,370).

The claim is drawn to a humanized monoclonal antibody that is free-end specific for the free n-terminus of an amyloid beta peptide binds to said free N-terminus said free terminus and does not bind to the amyloid beta-precursor protein from which said amyloid beta peptide may be proteolytically derived wherein the amyloid beta peptide is soluble in cerebrospinal fluid. The claim is also drawn to a neutralizing antibody that binds soluble amyloid beta.

Saido et al teach a polyclonal antibody 9204, that was produced using a synthetic hexamer peptide DAEFRC (Asp-Ala-Glu-Phe-Arg-Cys) conjugated to keyhole limpet hemocyanin. The antibody distinguished the fragments possessing the exact amino terminus of AB from the intact precursors and other fragments including the secretase products. Antibody 9204 also recognized synthetic AB1-40 peptide but not AB2-40 peptide. Furthermore, Saido et al teaches that binding of antibody 9204 to AFF-C100 was inhibited by the haptenic peptide DAEFRC, but not by MADEFTC or by AEFRHC. Saido et al teaches that this indicates that the antibody has strict specificity toward the cleavage site with an accuracy of 1 amino acid residue (i.e. the instant free-end specific N-terminal specific). Saido et al teaches that the use of the cleavage site specific antibody provides for better relative quantitiveness. (see page 15254-55, column 1, Results, first and second paragraphs). Saido et al teaches that "similar approaches for producing the proteolytic product specific antibodies will be applicable to resolving the differential carboxyl-terminal processing of AB peptides...". Saido et al differs by not teaching a monoclonal antibody with the properties of polyclonal antibody 9204.

Takeda teaches that monoclonal antibodies that are specific for the N-terminal and C-terminal of AB are useful for the detection of AB1-40 and AB1-42 for the detection of AB species *in vitro* (see page 5) Takeda teaches that AB1-40 is water soluble (page 4, lines 33-41). Takeda teach the N and C-terminal peptide sequence of AB1-40.

Vigo-Pelfrey et al teach the specific structures of beta amyloid peptides from human cerebrospinal fluid (CSF). Vigo-Pelfrey et al teach that amino acid sequencing reveals species of amyloid beta with N-termini of Asp1, Glu3, His6, Glu11 and Val12. Laser desorption mass spectrometry confirmed the presence of amyloid beta species containing 27, 28, 30, 34, 35, 40, 42 and 43 amino acids all beginning at Asp1. Vigo-Pelfrey et al is seen to teach the soluble amyloid beta species present in human CSF. Vigo-Pelfrey et al is seen to teach the free end C-terminus terminus of AB1-40 is present and soluble in CSF.

Goding teaches routine methods of making monoclonal antibodies with defined immunogens.

Queen teaches the production of chimeric antibodies (see column 11, lines 55-67) and chimeric humanized antibodies (see column 11, line 1-column 12, lines 4). Queen teach that the humanized antibodies can be used for diagnostic purpose and may either be labeled or unlabeled. A wide variety of labels can be employed such as radionuclides, fluors, enzymes etc. (column 20, lines 21-31) in a variety of immunoassays known to the art.

It would have been *prima facie* obvious to one having ordinary skill in the art at the time that the invention was made to use the teachings of Saido et al to generate humanized monoclonal antibodies free-end N-terminal specific antibodies that do not bind the precursor and bind species of soluble amyloid beta found in human cerebrospinal fluid using the conventional techniques of Queen et al and Goding et al because of the well established advantages of high-affinity, high specificity and unlimited supply that are central to monoclonal antibodies and Queen et al teach that humanized chimeric monoclonal antibodies can be used for diagnostic purposes. One would have been motivated to make monoclonal antibodies to decrease the lot to lot variability that can happen with polyclonal antisera and Takeda et al teach that the monoclonal antibodies are useful for the detection of AB1-40 and AB1-42 for the detection of AB species *in vitro* and that AB1-40 is water soluble and present in the human cerebrospinal fluid. One of ordinary skill in the art would have a reasonable expectation of success given the

demonstrated immunogenicity of the epitope. With respect to neutralizing antibody claims, the soluble form of amyloid beta is known in the art to be non-toxic (i.e. not neurotoxic) and as such any antibody that binds has neutralizing function since the antibody binds a non-toxic form of amyloid (i.e. soluble).

Claims 14, 33, 39 and new claim 54 are rejected under 35 U.S.C. 103(a) as being unpatentable over Takeda Chemical Industries Ltd., (EP 0 683 234 A1, published November 22, 1995, reference AC on PTOL-1449 filed 12 October 1999) in view of Saido et al (The Journal of Biochemistry, 269(21):15253-15257, 1994, Fee-based IDS Nov 16, 2001; hereinafter Saido A), Saido et al (The Journal of Biological Chemistry, 268(33):25239-25243, 1993; herein after Saido B), Vigo-Pelfrey et al (Journal of Neurochemistry, 61:1965-1968, 1993), Goding (Monoclonal Antibodies, Academic Press Inc., London 1983, pages 56-97) and Queen et al (US Patent No, 6,180,370).

The claims are drawn to a humanized monoclonal antibody that is free-end specific for the free C-terminus of an amyloid beta peptide 1-40 wherein the antibody binds to said free C-terminus said free terminus and does not bind to the amyloid beta-precursor protein from which said amyloid beta peptide may be proteolytically derived wherein the amyloid beta peptide is soluble in cerebrospinal fluid.

Takeda teaches that monoclonal antibodies that are specific for the N-terminal and C-terminal of AB peptides are useful for the detection of AB1-40 and AB1-42 for the detection of AB species *in vitro* (see pages 4-5). Takeda teaches that AB1-40 is water soluble (page 4, lines 33-41). Takeda teach the N and C-terminal peptide sequence of AB1-40. Takeda teach the monoclonal antibody BA-27a, that was considered to be specific for the C-terminus of beta amyloid (1-40), and weakly cross-reacted to beta-amyloid (1-38), (1-39) and beta amyloid (1-42) with a cross reactivity with 2% or less (page 34, lines 41-46). Takeda et al differs by not teaching a monoclonal antibody that has no cross-

reactivity as "uniquely recognizes" the free C-terminal of AB1-40 and does not recognize the precursor.

Saido A teaches a polyclonal antibody 9204, that was produced using a synthetic hexamer peptide DAEFRC (Asp-Ala-Glu-Phe-Arg-Cys) conjugated to keyhole limpet hemocyanin for the N-terminal of AB1-40. The antibody distinguished the fragments possessing the exact amino terminus of AB from the intact precursors and other fragments including the secretase products. Antibody 9204 also recognized synthetic AB1-40 peptide but not AB2-40 peptide. Furthermore, Saido et al teaches that binding of antibody 9204 to AFF-C100 was inhibited by the haptenic peptide DAEFRC, but not by MADEFTC or by AEFRHC. Saido et al teaches that this indicates that the antibody has strict specificity toward the cleavage site with an accuracy of 1 amino acid residue (i.e. the instant free-end specific N-terminal specific). Saido et al teaches that the use of the cleavage site specific antibody provides for better relative quantitiveness. (see page 15254-55, column 1, Results, first and second paragraphs). Saido et al teaches that "similar approaches for producing the proteolytic product specific antibodies will be applicable to resolving the differential carboxyl-terminal processing of AB peptides...". Saido et al teach that their unique methodology for producing such proteolytic produce-specific antibodies now seems to have general applicability (page 15254 (column 1, see first paragraph results section).

Saido B teaches a general technique for producing antibodies that specifically distinguish a proteolyzed form from a given intact form and are free-end specific.

Vigo-Pelfrey et al teach the specific structures of beta amyloid peptides from human cerebrospinal fluid (CSF). Vigo-Pelfrey et al teach that amino acid sequencing reveals species of amyloid beta with N-termini of Asp1, Glu3, His6, Glu11 and Val12. Laser desorption mass spectrometry confirmed the presence of amyloid beta species containing 27, 28, 30, 34, 35, 40, 42 and 43 amino acids all beginning at Asp1. Vigo-Pelfrey et al is

seen to teach the soluble amyloid beta species present in human CSF. Vigo-Pelfrey et al is seen to teach the free end C-terminus terminus of AB1-40 is present and soluble in CSF.

Goding teaches routine methods of making monoclonal antibodies with defined immunogens.

Queen teaches the production of chimeric antibodies (see column 11, lines 55-67) and chimeric humanized antibodies (see column 11, line 1-column 12, lines 4). Queen teach that the humanized antibodies can be used for diagnostic purpose and may either be labeled or unlabeled. A wide variety of labels can be employed such as radionuclides, fluors, enzymes etc. (column 20, lines 21-31) in a variety of immunoassays known to the art.

It would have been *prima facie* obvious to one having ordinary skill in the art at the time that the invention was made to use the teachings of the Saido A and B and Queen et al to generate free-end C-terminal specific AB1-40 humanized monoclonal antibodies that do not bind the precursor using the conventional end-peptide immunization techniques of Saido A and B combined with monoclonal antibody technology of Goding et al because of the well established advantages of high-affinity, high specificity and unlimited supply that are central to monoclonal antibodies and Takeda et al teach that antibodies with high sensitivity and specificity for amyloid peptide, including AB1-40 are desired and Vigo-Pelfrey teaches that AB1-40 species are soluble in CSF. One would have been motivated to make screen for free-end specific humanized monoclonal antibodies to eliminate the residual cross-reactivity of the monoclonal antibody BA-27(a) of Takeda and because Takeda et al teach that the prior art assays lack sensitivity and specificity and that highly specific monoclonal antibodies are useful for the detection of AB1-40 and AB1-42 species *in vitro* and that unique antibodies would reduce the background and increase the sensitivity of the immunoassay for AB1-40 and Queen et al teach that humanized monoclonal antibodies are useful for diagnostics. One of ordinary skill in the art would have a reasonable expectation of success given the demonstrated immunogenicity of the C-terminal epitope of AB1-40 as shown by Takeda and the success of Saido A for the N-

terminal epitope and that Saido A teaches that similar approaches will be applicable to resolving the differential carboxy terminal processing of AB peptides. With respect to neutralizing antibody claims, the soluble form of amyloid beta is known in the art to be non-toxic (i.e. not neurotoxic) and as such any antibody that binds has neutralizing function since the antibody binds a non-toxic form of amyloid (i.e. soluble).

Claims 23, 35 and 40 stand rejected under 35 U.S.C. 103(a) as being unpatentable over Saido et al (The Journal of Biochemistry, 269(21):15253-15257, 1994, Fee IDS Nov 16, 2001), Takeda Chemical Industries Ltd., (EP 0 683 234 A1, published November 22, 1995, reference AC on PTOL-1449 filed 12 October 1999), Vigo-Pelfrey et al (Journal of Neurochemistry, 61:1965-1968, 1993), Goding (Monoclonal Antibodies, Academic Press Inc., London 1983, pages 56-97) and Queen et al (US Patent No. 6,180,370) as applied to claim 14 and 24 above and further in view of Seubert et al (U.S. Patent 6,114,133, issued September 5, 2000 and filed November 14, 1994) and Duenas et al (BioTechniques, 16(3):476-483, 1994)

The claims are drawn to single chain antibodies that are free-N-terminal specific for AB peptide soluble in cerebrospinal fluid.

The teachings for Saido et al (The Journal of Biochemistry, 269(21):15253-15257, 1994, Fee-based IDS Nov 16, 2001) in view of Takeda Chemical Industries Ltd., (EP 0 683 234 A1, published November 22, 1995, reference AC on PTOL-1449 filed 12 October 1999), Vigo-Pelfrey et al (Journal of Neurochemistry, 61:1965-1968, 1993), Goding (Monoclonal Antibodies, Academic Press Inc., London 1983, pages 56-97) and Queen et al (US Patent No. 6,180,370) as combined are set forth supra. The references as combined fail to teach single chain antibodies.

Seubert et al teaches the use of antibodies that bind AB peptides in *in vitro* or *in vivo* assays that screen for inhibitors of AB peptide formation (see columns 4-5, Summary of the Invention). Seubert et al teach that in addition to monoclonal antibodies, "... the

detection techniques of the present invention will also be able to use antibody fragments, such as F(ab), Fv, VL, VH, and other fragments." Seubert et al also teach that "It would also be possible to employ recombinantly produced antibodies (immunoglobulins) and variation thereof as now well described in the patent and scientific literature. See, for example EPO 8430268.0; EPO 85102665.8; EPO 85305604.2; PCT/GB 85/00392; EPO 85115311.4; PCT/US 86/002269; and Japanese application 85239543." (see column 10, first full paragraph).

Duenas et al teach art accepted conventional methods of intra- and extracellular expression of a single chain Fv antibody fragment (scFv) in *E. coli*. Duenas et al teach that cloning of immunoglobulin variable regions and bacterial expression of antibody fragments was routinely performed in the art at the time that this invention was made (see page 476, column 2, Introduction).

It would have been *prima facie* obvious to one having ordinary skill in the art at the time that the invention was made to modify the free-end, N-terminal specific humanized monoclonal antibody according to the combination of Saido et al (The Journal of Biochemistry, 269(21):15253-15257, 1994, Fee-based IDS Nov 16, 2001) in view of Takeda Chemical Industries Ltd., (EP 0 683 234 A1, published November 22, 1995, reference AC on PTOL-1449 filed 12 October 1999), Goding (Monoclonal Antibodies, Academic Press Inc., London 1983, pages 56-97) and Queen et al (US Patent No, 6,180,370).
supra, by means of expression as a single chain Fv antibody fragment (scFv) according to the vectors and methodology of Duenas et al because Seubert et al teach that Fv and other antibody fragments including those that have been recombinantly produce that bind AB peptides are useful in a variety of detection techniques for use in screening or diagnostic assays. With respect to neutralizing antibody claims, the soluble form of amyloid beta is known in the art to be non-toxic (i.e. not neurotoxic) and as such any antibody that binds has neutralizing function since the antibody binds a non-toxic form of amyloid (i.e. soluble).

Claims 23, 38 and 40 stand rejected under 35 U.S.C. 103(a) as being unpatentable over Takeda Chemical Industries Ltd., (EP 0 683 234 A1, published November 22, 1995, reference AC on PTOL-1449 filed 12 October 1999), Saido et al (The Journal of Biochemistry, 269(21):15253-15257, 1994, Fee-based IDS Nov 16, 2001; hereinafter Saido A), Saido et al (The Journal of Biological Chemistry, 268(33):25239-25243, 1993; herein after Saido B), Vigo-Pelfrey et al (Journal of Neurochemistry, 61:1965-1968, 1993), Goding (Monoclonal Antibodies, Academic Press Inc., London 1983, pages 56-97) and Queen et al (US Patent No, 6,180,370) as applied to claims 14, 33, 39 and 54 above and further in view of Seubert et al (U.S. Patent 6,114,133, issued September 5, 2000 and filed November 14, 1994) and Duenas et al (BioTechniques, 16(3):476-483, 1994).

The teachings of Takeda Chemical Industries Ltd., (EP 0 683 234 A1, published November 22, 1995, reference AC on PTOL-1449 filed 12 October 1999) in view of Saido et al (The Journal of Biochemistry, 269(21):15253-15257, 1994, Fee-based IDS Nov 16, 2001; hereinafter Saido A), Saido et al (The Journal of Biological Chemistry, 268(33):25239-25243, 1993; herein after Saido B), Vigo-Pelfrey et al (Journal of Neurochemistry, 61:1965-1968, 1993), Goding (Monoclonal Antibodies, Academic Press Inc., London 1983, pages 56-97) and Queen et al (US Patent No, 6,180,370) are set forth above. The references as combined differ by not teaching single chain antibodies.

Seubert et al teaches the use of antibodies that bind AB peptides in *in vitro* or *in vivo* assays that screen for inhibitors of AB peptide formation (see columns 4-5, Summary of the Invention). Seubert et al teach that in addition to monoclonal antibodies, "... the detection techniques of the present invention will also be able to use antibody fragments, such as F(ab), Fv, VL, VH, and other fragments." Seubert et al also teach that "It would also be possible to employ recombinantly produced antibodies (immunoglobulins) and variation thereof as now well described in the patent and scientific literature. See, for example EPO 8430268.0; EPO 85102665.8; EPO 85305604.2; PCT/GB 85/00392; EPO

85115311.4; PCT/US 86/002269; and Japanese application 85239543." (see column 10, first full paragraph).

Duenas et al teach art accepted conventional methods of intra- and extracellular expression of a single chain Fv antibody fragment (scFv) in *E. coli*. Duenas et al teach that cloning of immunoglobulin variable regions and bacterial expression of antibody fragments was routinely performed in the art at the time that this invention was made (see page 476, column 2, Introduction).

It would have been *prima facie* obvious to one having ordinary skill in the art at the time that the invention was made to modify the free-end, C-terminal specific humanized monoclonal antibody that binds a soluble CSF amyloid beta peptide according to the combination of Takeda Chemical Industries Ltd., (EP 0 683 234 A1, published November 22, 1995, reference AC on PTOL-1449 filed 12 October 1999) in view of Saido et al (The Journal of Biochemistry, 269(21):15253-15257, 1994, Fee-based IDS Nov 16, 2001; hereinafter Saido A), Saido et al (The Journal of Biological Chemistry, 268(33):25239-25243, 1993; herein after Saido B), Vigo-Pelfrey et al (Journal of Neurochemistry, 61:1965-1968, 1993) and Goding (Monoclonal Antibodies, Academic Press Inc., London 1983, pages 56-97) above, by means of expression as a single chain Fv antibody fragment (scFv) according to the vectors and methodology of Duenas et al because Seubert et al teach that Fv and other antibody fragments including those that have been recombinantly produce that bind AB peptides are useful in a variety of detection techniques for use in screening or diagnostic assays. With respect to neutralizing antibody claims, the soluble form of amyloid beta is known in the art to be non-toxic (i.e. not neurotoxic) and as such any antibody that binds has neutralizing function since the antibody binds a non-toxic form of amyloid (i.e. soluble).

Applicants argue since the other rejections fail for reasons previously presented,

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then these rejections would also fail. This is not persuasive because the other rejections do not fail for reasons set forth in all the previous office actions of record, motivation has been specifically and explicitly provided and all limitations have been met for reasons set forth therein. Further, *In re Fine*, 837 F.2d 1071, 1075, 5U.S.P.Q.2d 1959 (Fed. Cir. 1988) states that under section 103 a *prima facie* case of obviousness can be established by showing some objective teaching in the prior art ***or that knowledge generally available to one of ordinary skill in the art can lead the individual to combine the references.*** See also *In re Jones*, 958 F.2d 347, 21 USPQ2d 1941 (Fed. Cir. 1992). Furthermore, the courts have held "The test of obviousness is not express suggestion of the claimed invention in any or all of the references but rather what the references taken collectively would suggest to those of ordinary skill in the art presumed to be familiar with them." See *In re Rosselet*, 146 USPQ 183, 186 (CCPA 1965). "There is no requirement (under 35 USC 103(a)) that the prior art contain an express suggestion to combine known elements to achieve the claimed invention. Rather, the suggestion to combine may come from the prior art, as filtered through the knowledge of one skilled in the art." *Motorola, Inc. v. Interdigital Tech. Corp.*, 43 USPQ2d 1481, 1489 (Fed. Cir. 1997). Finally, an obviousness determination is not the result of a rigid formula disassociated from the consideration of the facts of a case. Indeed, the common sense of those skilled in the art demonstrates why some combinations would have been obvious where others would not. See *KSR Int'l Co. v. Teleflex Inc.*, 550 U.S. , 2007 U.S. LEXIS 4745, 2007 WL 1237837, at *12 (2007) ("The combination of familiar elements according to known methods is likely to be obvious when it does no more than yield predictable results."). This composition is merely a combination of known elements, yielding predictable results. Therefore, none of Applicants arguments are persuasive.

New Rejections Based on Amendment

Claim Rejections - 35 USC § 112

The following is a quotation of the second paragraph of 35 U.S.C. 112:

The specification shall conclude with one or more claims particularly pointing out and distinctly claiming the subject matter which the applicant regards as his invention.

Claims 50 and 54 are rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

The claims recite a range of amino acid residues of amyloid B-peptide. However, it is noted that specific structures of beta amyloid peptides from human cerebrospinal fluid (CSF). Vigo-Pelfrey et al teach that amino acid sequencing reveals species of amyloid beta with N-termini of Asp1, Glu3, His6, Glu11 and Val12. As such, the claims recite a range of residues within a genus of beta amyloid peptides as such, the skilled artisan would not be readily apprised of the residues to which the antibody binds.

Status of Claims

Claims 14, 23, 24, 33, 35, 38-40, 50 and 54 are rejected. Claims 41-49, 51-53 and 55-57 are withdrawn from consideration.

Conclusion

Applicant's amendment necessitated the new ground(s) of rejection presented in this Office action. Accordingly, **THIS ACTION IS MADE FINAL**. See MPEP § 706.07(a). Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire **THREE MONTHS** from the mailing date of this action. In the event a first reply is filed within **TWO MONTHS** of the mailing date of this final action and the advisory action is not mailed until after the end of the **THREE-MONTH** shortened statutory period, then the

shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than SIX MONTHS from the date of this final action.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Patricia A. Duffy whose telephone number is 571-272-0855. The examiner can generally be reached on M-Th 7:30 am - 6:00 pm. If attempts to reach the examiner by telephone are unsuccessful, the examiner's Supervisors, Robert Mondesi can be reached at 571-272-0956.

The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

/Patricia A. Duffy/
Primary Examiner